A Hydroalcoholic Extract from the Leaves of Nerium oleander Inhibits Glycolysis and Induces Selective Killing of Lung Cancer Cells

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Introduction
Cardiac glycosides are a group of natural products that share a steroid-like structure with an unsaturated lactone ring and the ability to inhibit the Na+/K+-ATPase pump. Numerous cardiac glycosides (e.g., digoxin, ouabain, oleandrin, and proscillaridin) have been isolated from plants (e.g., Digitalis purpurea, Digitalis lanata, Strophantus gratissus, Nerium oleander, and Urginea maritima). Several cardiac glycosides have also been found in amphibians and mammals, including digoxin, oua-
bain, bufalin, marinobufagenin, and telecinobufagin. Some cardiac glycosides are used in cardiology for the treatment of cardiac congestion and some types of cardiac arrhythmias. The mechanism by which these drugs affect cardiac contractility is known to be mediated by a highly specific inhibition of Na⁺/K⁺-ATPase [1–3].

Over the years, there have been a variety of periodic reports suggesting that cardiac glycosides may have an anticancer utilization (reviewed in [4–8]). In vitro and ex vivo experiments have revealed that some cardiac glycosides (e.g., digitoxin) induce selective anticancer effects [4,9,10], which may occur at concentrations commonly found in the plasma of patients treated with these drugs [11]. Some cardiac glycosides have also shown potent and selective anticancer effects in mice harboring human malignant cells [12,13]; however, these results should be interpreted cautiously, as mouse cells are much more resistant than human cells to the cytotoxic effects of cardiac glycosides, and it is not clear whether such selectivity is due to selective inhibition of tumor cells or to interspecies differences in sensitivity [14]. The cardiac drugs digitoxin and digoxin, the semisynthetic cardiac glycoside UNBS1450, and two extracts from Nerium oleander Apocynaceae have entered clinical trials for the treatment of cancer (see http://clinicaltrials.gov/ and ref. [6,7,15,16]). Recently, however, Perne et al. reported results suggesting that cardiac glycoside-induced cytotoxicity was mediated by general protein synthesis inhibition and was not selective for cancer cells, raising concerns about ongoing clinical trials testing cardiac glycoside-induced cytotoxicity as anticancer agents [17]. Later, Hallbook et al. observed that some cardiac glycosides, particularly digitoxin, induced ex vivo selective anticancer effects in leukemia cells and found that protein synthesis inhibition by cardiac glycosides at concentrations corresponding to IC₅₀ values did not occur in all types of cancer cells [10]. These data suggest that cardiac glycosides may induce selective anticancer effects only in some types of cancer.

Several mechanisms of action have been proposed to participate in the cytotoxic activity of cardiac glycosides (reviewed in [5–8,16]). However, it is unclear why cancer cells are generally more susceptible than nonmalignant cells to the cytotoxic activity of these compounds. Recent data have revealed that cancer cells have a higher reliance on glycolysis for their survival than normal cells, and that the inhibition of glycolysis may cause selective anticancer effects [5,18,19]. In this communication, we have assessed the selective cytotoxic activity of a hydroalcoholic extract from the leaves of the cardenolide-containing plant Nerium oleander against A549 lung cancer cells vs. MRC5 nonmalignant lung fibroblasts and have evaluated possible mechanisms involved in this activity. Results show that this extract induces selective killing of lung cancer cells and a marked inhibition of glycolysis.

Materials and Methods

Plant material, preparation of the extract, and determination of cardiac glycosides content

The leaves of Nerium oleander L. were collected in June 2010 in Seville (Spain; 37°22′16″N, 5°59′27″W). A voucher specimen (278048) was authenticated by Dr. F. García and is deposited in the herbarium at the Department of Vegetal Biology and Ecology, Faculty of Biology, University of Seville. Fresh leaves of N. oleander were extracted with ethanol: water (1:1) at 60°C for 1 hour by using an ultrasound water bath apparatus. Ethanol was eliminated in a rotary vacuum evaporator, and the remaining water solution was lyophilized, with an extraction yield of 2.3%. The cardiac glycoside content of N. oleander was determined with the Kedde reaction, a colorimetric technique that allows the determination of unsaturated penacyclic lactones (present in cardiac glycosides from N. oleander) by using 3,5-dinitrobenzoic acid [20]. Briefly, a 3% solution of 3,5-dinitrobenzoic acid in ethanol was mixed in the ratio 1:1 with a solution of 2 M NaOH in distilled water. 100 µL of this mixture was mixed with 150 µL of NOE or 2-furanone in ethanol at different concentrations. Optical densities were measured at 540 nm on a multiwell plate spectrophotometer reader. Based on the standard 2-furanone (lactone of cardenolides), the percentage of cardenolides in the extract was determined, and was expressed as the mean ± SEM.

Chemicals and cell lines

Cisplatin (99.9%), dichloroacetate (98%), NAC (99%), 2-furanone (97%), and catalase were purchased from Sigma. The human A549 lung cancer cell line, the human embryo lung fibroblastic MRC-5 cell line and the human HT29 colon adenocarcinoma cell line were purchased from European Collection of Cell Cultures. The human UACC-62 melanoma cell line was purchased from American Type Culture Collection. The HR-deficient V8 cell line (V79 Chinese hamster lung cells mutated in BRCA2) and the VCB82 cell line (V8 cells complemented with human BRCA2) were kindly provided by Dr. Helleday. All cell lines were maintained in DMEM supplemented with 2 mM glutamine, 50 µg/mL penicillin, 50 µg/mL streptomycin, and 10% fetal bovine serum, and were cultured at 37°C in a humidified atmosphere containing 5% CO₂. Cell culture reagents were obtained from Life Technologies.

Assay for cytotoxic activity

The MTT assay is a colorimetric technique that allows the quantitative determination of cell viability. It is based on the capability of viable cells to transform the MTT salt (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) into a formazan dye. Exponentially growing cells were seeded into 96-well plates and drugs were added 24 h later. Following the incubation period specified in the figures and table legends, the medium was removed, and 125 µL MTT (1 mg/mL in medium) was added to each well for 4 hours. Then, 80 µL 20% SDS in 0.02 M HCl were added, plates were incubated for 10 hours at 37°C, and optical densities were measured at 540 nm on a multiwell plate spectrophotometer reader. Cell viability was expressed as percentage in relation to controls. All data were averaged from at least three independent experiments and were expressed as means ± SEM.

Immunofluorescence γ-H2AX focus assay

The immunofluorescence γ-H2AX focus assay is a sensitive technique to evaluate DNA damage. It is based on the ability of DSBs to trigger phosphorylation of histone H2AX on Ser-139, which leads to the formation of nuclear foci that can be visualized with anti-γH2AX antibodies [21,22]. Cells were seeded on coverslips and allowed to attach for 24 hours. After treatments, cells were washed three times with PBS, fixed with 4% paraformaldehyde in PBS for 10 min at room temperature and washed again three times with PBS. After fixation, cells were permeabilized with 0.5% Triton X-100 in PBS for 5 minutes and then blocked three times with 0.1% Tween 20, 1% BSA in PBS for 5 minutes each. Cells were then incubated for 30 min with a mouse anti-γH2AX monoclonal antibody (Upstate, 1:1000 dilution). Cells were washed three times with PBS and blocked three times prior to the incuba-
tion with a secondary anti-mouse antibody linked to Alexa Fluor 488 (Invitrogen, 1:1000 dilution) for 1 h. Cells were washed with PBS, blocked and washed again with PBS as indicated before. DNA was stained with DAPI, and immunofluorescence was observed at 40-fold magnification with an Olympus BX 61 microscope. A total of ~200 cells/dose were scored, and cells with 20 or more foci were scored as positive. Ionizing radiation was used as a positive control for the assay; cells were exposed to 4 Gy of ionizing radiation using an X-ray irradiator (Philips MU15F) operated at 100 KV and a dose rate of 1 Gy/min [23].

Inhibition of glycolysis
Glycolysis inhibition was assessed by measuring concentrations of glucose (initial product of glycolysis) and lactate (final product of glycolysis) in control and treated cells. Briefly, 10^6 cells were exposed in 6-well plates to the tested compounds for 8 h, and glucose and lactate concentrations were determined in cell supernatants by using the Accutrend® Plus analyzer together with Accutrend glucose strips and BM-Lactate Strips (Roche Diagnostics). After calibrating the instrument with glucose and lactate calibration strips, test strips were used to determine glucose and lactate levels via colorimetric-oxidase mediator reactions according to the manufacturer’s instructions [24]. Results are expressed as percentage of lactate production and percentage of glucose consumption in relation to untreated cells and are shown as means ± SEM of three independent experiments.

Statistical analysis
All data were averaged from at least three independent experiments and were expressed as means ± SEM. For statistical analysis we used the t-test (paired, two-tailed). A p value < 0.05 is not considered statistically significant and is not represented by any symbol. A p value < 0.05 is considered to correspond with statistical significance and is indicated with an asterisk (*), a p value < 0.01 is indicated with a double asterisk (**), and a p value < 0.001 is indicated with a triple asterisk (***)

Results and Discussion

Extracts from the leaves of the cardiac glycoside-containing plant *Nerium oleander* L. have shown anticancer effects in preclinical studies and have entered phase I clinical trials [16, 25, 26]. Because recent data suggest that cardiac glycosides may induce selective anticancer effects only in some types of cancer cells [10, 17], the aim of this work was to evaluate whether an extract from this plant induced selective cytotoxic activity in lung cancer cells and to evaluate possible mechanisms involved in this activity. A hydroalcoholic extract from the leaves of *N. oleander* was prepared, and its content in cardiac glycosides (cardenolides) was determined to be 4.75 ± 0.32% with the Kedde reaction, which is an accepted methodology for standardizing cardenolides. Another reason for selecting the Kedde reaction for standardizing the extract is that, unlike a chromatographic profile of several of its major constituents, this reaction gives its total cardenolide content. Several extracts from *N. oleander* (but not any of their constituents) have entered clinical trials, therefore suggesting that their anticancer activity is mediated by their total cardenolide content rather than by any particular constituent of the extracts. In addition, it is much easier for any researcher wanting to reproduce our results to use the Kedde reaction than to carry out a chromatographic profile of several constituents of the extract selected by us.

A549 lung cancer cells and MRC5 nonmalignant lung fibroblasts were exposed for 48 h to several concentrations of this NOE under the same experimental conditions, and cell viability was estimated with the MTT assay. Results, represented in Fig. 1, show that NOE exhibited selective cytotoxicity against the A549 cancer cell line, which was comparable to that of the anticancer drug cisplatin. The IC_{50} value (means ± SEM) in this cancer cell line was approximately 10-fold lower for NOE (0.27 ± 0.04 µg/mL) than for cisplatin (3.51 ± 0.37 µg/mL; 11.67 ± 3.54 µM). Several compounds were screened against these two cell lines and showed no selective cytotoxicity [27, 28]. We have also prepared and assessed the cytotoxic activity of an extract of commercial broccoli under the same experimental conditions used for NOE, and we did not observe selective cytotoxicity against the cancer cell line (data not shown). We also evaluated the cytotoxic activity of NOE in the human HT29 colon cancer cell line and in the human UACC62 melanoma cell line and observed that these cancer cells were more resistant than A549 lung cancer cells to the cytotoxicity of NOE; the IC_{50} values (means ± SEM) were 3.95 ± 0.37 µg/mL in HT29 cells and 1.73 ± 0.07 µg/mL in UACC62 cells. Although NOE inhibited the growth of the cancer cell lines HT29 and UACC62 at low concentrations, these concentrations also inhibited the growth of the nonmalignant cell line MRC5 (Fig. 1B). Overall, these data indicate that NOE inhibits the growth of A549 cancer cells potently and selectively.

Our next goal was to evaluate possible mechanisms involved in the selective cytotoxicity of NOE. We initially evaluated whether the formation of ROS played a role in the cytotoxicity of the extract. Accumulating data suggest that cancer cells have higher basal levels of ROS than nonmalignant cells, and that the induction of a specific increase in ROS levels by pro-oxidant agents may lead to cytotoxic concentrations in cancer cells but not in normal cells. Indeed, oxidative stress has been shown to play an important role in the anticancer activity of several chemotherapeutic agents commonly used in cancer treatment [29–31] and in the cytotoxicity of many natural products [32], including that of the major constituent of *N. oleander*, oleandrin [33]. A549 cells were treated with NOE in the presence and absence of the antioxidants catalase and N-acetylcysteine, and cell viability was estimated with the MTT assay. Both antioxidants slightly but significantly prevented the cytotoxic activity of NOE (Fig. 2), therefore suggesting that ROS formation participates in the cytotoxicity of NOE but does not play a major role.

DNA-damaging compounds may induce selective cytotoxicity towards tumor cells. These cells commonly have mutations in DNA repair genes which make them vulnerable to the cytotoxic activity of specific DNA-damaging compounds. Unlike tumor cells, normal proliferating cells have an intact DNA damage response that would allow them to repair the DNA damage and therefore to survive treatment with these DNA-damaging compounds [34–36]. Because previous reports showed that some cardiac glycosides induce DNA damage [11, 37–39], we tested whether NOE induced DNA damage and if such damage participated in the cytotoxicity of the extract. We initially tested the cytotoxicity of NOE in cells with and without BRCA2 (a tumor suppressor gene which plays a critical role in DNA damage repair via the HR repair pathway). The HR-deficient VCB cell line (V79 Chinese hamster lung cells mutated in BRCA2) and the VCB82 cell line (VCB cells complemented with human BRCA2) were exposed for 48 h to several concentrations of NOE, and cell viability was then estimated with the MTT assay. Results, represented in Fig. 3A, show that the cytotoxic activity of NOE in cells lacking BRCA2.
(deficient in HR repair) was slightly but significantly higher than that in parental cells, therefore suggesting that NOE can induce DNA damage and that NOE-induced DNA damage may play a role in its cytotoxicity. It is important to note that the cytotoxicity of NOE in these rodent cells was over 100-fold lower than that in human cells (Fig. 1), which is highly specific for cardiac glycosides [40]. We next used the immunofluorescence γ-H2AX focus assay to directly measure the levels of DNA damage induced by NOE in both cell lines and detected moderate levels of DNA damage in cells exposed for 24 h to NOE 320 µg/mL (Fig. 3B). As expected, the levels of DNA damage in cells lacking BRCA2 (deficient in HR repair) were somewhat higher than those in parental cells. Taken as a whole, these data suggest that NOE induces DNA damage and that this DNA damage participates in its cytotoxic activity but does not play a major role.

It has been known for some time that glycolysis is coupled to sodium and potassium transport processes and that some cardiac glycosides (e.g., ouabain) can inhibit glycolysis in a variety of non-malignant cells [41, 42]. It has recently been found that cancer cells are more reliant on glycolysis for their survival than non-malignant cells (reviewed in [18,31]). Based on these findings, we previously proposed that cardiac glycosides might inhibit glycolysis in cancer cells and that such inhibition might play a critical role in the selective cytotoxicity of some cardiac glycosides (e.g., digitoxin) towards cancer cells [5]. To test if NOE could inhibit glycolysis, A549 cells were exposed to NOE for 8 h, and glucose and lactate concentrations were determined in cell supernatants. Results, represented in Fig. 4, show that A549 cells exposed to NOE (1 and 10 µg/mL) exhibited a marked reduction in glucose consumption and lactate production, comparable to those observed in cells exposed to the glycolysis inhibitor DCA 32 mM. No significant changes in glucose consumption and lactate production were observed in cells exposed to cisplatin 32 µM.

would decrease glycolysis activity by inhibition of the key glycolytic enzyme phosphofructokinase (discussed in reference [5]). In addition, inhibition of the Na+/K+-ATPase pump may restrict the activity of SGLTs, which couple glucose entry into some types of cells with the activity of this pump [5]. Although they are typically found in small intestine and renal epithelial cells, clinical data have revealed that these transporters (SGLT2) are overexpressed in lung cancer cells that have metastasized to other organs [43]. This suggests that SGLT2 plays a role in glucose uptake in the metastatic lesions of lung cancer, and that inhibition of the Na+/K+-ATPase pump by cardiac glycosides may inhibit glycolysis by reducing glucose entry into the cells. The inhibition of glycolysis induced by NOE in A549 lung cancer cells may therefore be caused by inhibition of the Na+/K+-ATPase pump, which would lead to both the inhibition of the glycolytic enzyme PFK and the inhibition of glucose entry into the cells.

Glycolysis inhibition is well known to produce cytotoxicity. The main cellular roles of glycolysis are to provide building blocks for biosynthesis and ATP, and evidence suggests that inhibition of these processes may have a higher impact on the viability of cancer cells than on that of nonmalignant cells. Indeed, glycolysis inhibition has been proposed to be an anticancer strategy to selectively kill cancer cells [18, 31]. Although here we report that NOE induces selective cytotoxicity against lung cancer cells and...
that this extract induces a marked inhibition of glycolysis, we cannot conclude that NOE-induced inhibition of glycolysis is responsible for its selective anticancer activity. Evidence suggests that the expression and cellular location of Na\(^+/\)K\(^{-}\)-ATPase alpha subunits in different types of cells (i.e., rodent cells, human cancer cells, and human nonmalignant cells) may explain why different cells are more or less susceptible to the cytotoxic activity of cardiac glycosides [44–46]. The study of the possible association between the expression and cellular location of Na\(^+/\)K\(^{-}\)-ATPase alpha subunits and the ability of cardiac glycosides to inhibit glycolysis would help reveal whether the inhibition of glycolysis by these compounds plays a role in their selective cytotoxicity. Since most cancer chemotherapy regimens include a combination of drugs, and since platinum compounds are widely used in the treatment of lung cancer, we assessed the cytotoxic activity of NOE in combination with cisplatin in A549 lung cancer cells. Then, we calculated the parameter CI with the computer software Compusyn; this parameter is based on the Chou-Talalay method [47]. A549 cells were exposed for 44–48 h to several concentrations of NOE, cisplatin, and NOE in combination with cisplatin (NOE was added 4 h before cisplatin or vice versa). Cell viability was assessed by the MTT assay. The parameter combination index (CI) was calculated with the computer software Compusyn. A CI value < 0.9 is considered to be synergism and is represented by +++ for very strong synergism (CI < 0.1), ++ for strong synergism (CI = 0.1–0.3), + for synergism (CI = 0.3–0.7), * for moderate synergism (CI = 0.7–0.85), and * for slight synergism (CI = 0.85–0.9). A CI value between 0.9 and 1.1 corresponds with the additive effect and is indicated with *. A CI value > 1.1 is considered to be antagonism and is represented by – for slight antagonism (CI = 1.1–1.2), –* for moderate antagonism (CI = 1.2–1.45), –** for antagonism (CI = 1.45–3.3), –*** for strong antagonism (CI = 3.3–10), and –**** for very strong antagonism (CI > 10).

## Table 1: Cytotoxic activity of NOE in combination with the anticancer drug cisplatin.

<table>
<thead>
<tr>
<th>First compound added</th>
<th>% Cell viability</th>
<th>Second compound added</th>
<th>% Cell viability</th>
<th>Combination</th>
<th>% Cell viability</th>
<th>CI</th>
<th>Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Concentration</td>
<td></td>
<td>Concentration</td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1 µM cisplatin</td>
<td>85.97 ± 1.63</td>
<td>32 ng/ml NOE</td>
<td>81.30 ± 2.07</td>
<td>80.11 ± 2.44</td>
<td>1.40</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>100 ng/ml NOE</td>
<td>65.57 ± 0.24</td>
<td>320 ng/ml NOE</td>
<td>28.50 ± 4.01</td>
<td>24.69 ± 1.52</td>
<td>0.76</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>320 ng/ml NOE</td>
<td>28.50 ± 4.01</td>
<td>320 ng/ml NOE</td>
<td>28.50 ± 4.01</td>
<td>22.56 ± 1.61</td>
<td>0.71</td>
<td>++</td>
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</tr>
<tr>
<td>3 µM cisplatin</td>
<td>72.59 ± 2.81</td>
<td>32 ng/ml NOE</td>
<td>81.30 ± 2.07</td>
<td>49.81 ± 0.32</td>
<td>0.94</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>10 µM cisplatin</td>
<td>53.82 ± 0.20</td>
<td>100 ng/ml NOE</td>
<td>65.57 ± 0.24</td>
<td>37.15 ± 1.48</td>
<td>0.81</td>
<td>++</td>
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<tr>
<td>32 µM cisplatin</td>
<td>33.00 ± 2.56</td>
<td>100 ng/ml NOE</td>
<td>65.57 ± 0.24</td>
<td>24.20 ± 2.84</td>
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<td>320 ng/ml NOE</td>
<td>28.50 ± 4.01</td>
<td>320 ng/ml NOE</td>
<td>28.50 ± 4.01</td>
<td>13.82 ± 0.52</td>
<td>0.58</td>
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<tr>
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<td>84.91 ± 3.92</td>
<td>1.20</td>
<td>–</td>
<td>–</td>
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<tr>
<td>100 ng/ml NOE</td>
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<td>320 ng/ml NOE</td>
<td>24.62 ± 5.31</td>
<td>64.32 ± 4.00</td>
<td>1.00</td>
<td>±</td>
<td></td>
</tr>
<tr>
<td>320 ng/ml NOE</td>
<td>24.62 ± 5.31</td>
<td>32 ng/ml NOE</td>
<td>90.15 ± 2.17</td>
<td>29.10 ± 3.90</td>
<td>1.09</td>
<td>±</td>
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<tr>
<td>100 ng/ml NOE</td>
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<td>320 ng/ml NOE</td>
<td>24.62 ± 5.31</td>
<td>63.03 ± 1.93</td>
<td>1.17</td>
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<tr>
<td>320 ng/ml NOE</td>
<td>24.62 ± 5.31</td>
<td>32 ng/ml NOE</td>
<td>90.15 ± 2.17</td>
<td>34.03 ± 3.62</td>
<td>1.31</td>
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<td>10 µM cisplatin</td>
<td>59.78 ± 2.40</td>
<td>53.93 ± 3.62</td>
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<td>±</td>
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<td>–</td>
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<td>100 ng/ml NOE</td>
<td>72.90 ± 5.12</td>
<td>28.95 ± 0.12</td>
<td>1.20</td>
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<tr>
<td>32 ng/ml NOE</td>
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<td>320 ng/ml NOE</td>
<td>28.50 ± 4.01</td>
<td>39.37 ± 0.93</td>
<td>0.98</td>
<td>±</td>
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<tr>
<td>1 µM cisplatin</td>
<td>85.97 ± 1.63</td>
<td>32 ng/ml NOE</td>
<td>81.30 ± 2.07</td>
<td>36.72 ± 0.55</td>
<td>1.09</td>
<td>±</td>
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<tr>
<td>100 ng/ml NOE</td>
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<td>320 ng/ml NOE</td>
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<td>28.45 ± 0.90</td>
<td>1.47</td>
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</table>

A549 cancer cells were exposed for 44–48 h to several concentrations of NOE, cisplatin, and NOE in combination with cisplatin (NOE was added 4 h before cisplatin or vice versa). Cell viability was assessed by the MTT assay. The parameter combination index (CI) was calculated with the computer software Compusyn. A CI value < 0.9 is considered to be synergism and is represented by +++ for very strong synergism (CI < 0.1), ++ for strong synergism (CI = 0.1–0.3), + for synergism (CI = 0.3–0.7), * for moderate synergism (CI = 0.7–0.85), and * for slight synergism (CI = 0.85–0.9). A CI value between 0.9 and 1.1 corresponds with the additive effect and is indicated with *. A CI value > 1.1 is considered to be antagonism and is represented by – for slight antagonism (CI = 1.1–1.2), –* for moderate antagonism (CI = 1.2–1.45), –** for antagonism (CI = 1.45–3.3), –*** for strong antagonism (CI = 3.3–10), and –**** for very strong antagonism (CI > 10).

In conclusion, this work reports that an extract from the leaves of the cardiac glycoside-containing plant *Nerium oleander* exerts selective cytotoxic activity towards lung cancer cells and induces a marked inhibition of glycolysis that may play a role in this activity. It also shows that NOE-induced DNA damage and ROS formation participate moderately in its cytotoxicity, and that the administration of NOE after the anticancer drug cisplatin may induce synergistic cytotoxicity. Since phase I clinical trials have revealed that extracts from the leaves of *Nerium oleander* are well tolerated and may induce anticancer effects [16, 25, 26], the present results support their possible advancement into phase II clinical trials for the treatment of lung cancer.

## Conflict of Interest

The authors declare that they do not have conflicts of interest.

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